

MICROBIAL AND PHYSIOCHEMICAL ANALYSIS OF WATER FROM SEMENGGOK INLAND FISHERY CENTRE, KUCHING, SARAWAK

Loke Xin Ya (70241)

Bachelor of Science with Honours (Resource Biotechnology) 2022

MICROBIAL AND PHYSIOCHEMICAL ANALYSIS OF WATER FROM SEMENGGOK INLAND FISHERY CENTRE, KUCHING, SARAWAK

LOKE XIN YA

A thesis submitted in partial fulfilment of the requirement of the Degree Bachelor of Science with Honours (Resource Biotechnology)

SUPERVISOR: DR. LEE KUI SOON CO-SUPERVISOR: DR. SAMUEL LIHAN

Programme of Resource Biotechnology Faculty of Resource Science and Technology UNIVERSITI MALAYSIA SARAWAK 2022

UNIVERSITI MALAYSIA SARAWAK

Grade:

Please tick (√) Final Year Project Report Masters PhD

DECLARATION OF ORIGINAL WORK

This declaration is made on the 12th day of June 2022.

Student's Declaration:

I, LOKE XIN YA, 70241, FACULTY OF RESOURCE SCIENCE AND TECHNOLOGY, hereby declare that the work entitled, MICROBIAL AND PHYSIOCHEMICAL ANALYSIS OF WATER FROM SEMENGGOK INLAND FISHERY CENTRE, KUCHING, SARAWAK is my original work. I have not copied from any other students' work or from any other sources except where due reference or acknowledgement is made explicitly in the text, nor has any part been written for me by another person.

12.06.2022

Date submitted

LOKE XIN YA (70241) Name of the student (Matric No.)

Supervisor's Declaration:

Received for examination by: LE

LEE KUI SOON

Date: 14 July 2022

(Name of the supervisor)

I declare this Project/Thesis is classified as (Please tick $(\sqrt{})$):

CONFIDENTIAL (Contains confidential information under the Official Secret Act 1972)*
RESTRICTED (Contains restricted information as specified by the organisation where research was done)*

OPEN ACCESS

Validation of Project/Thesis

I therefore duly affirmed with free consent and willingness declared that this said Project/Thesis shall be placed officially in the Centre for Academic Information Services with the abide interest and rights as follows:

- This Project/Thesis is the sole legal property of Universiti Malaysia Sarawak (UNIMAS).
- The Centre for Academic Information Services has the lawful right to make copies for the purpose of academic and research only and not for other purpose.
- The Centre for Academic Information Services has the lawful right to digitise the content to for the Local Content Database.
- The Centre for Academic Information Services has the lawful right to make copies of the Project/Thesis for academic exchange between Higher Learning Institute.
- No dispute or any claim shall arise from the student himself / herself neither third party on this Project/Thesis once it becomes sole property of UNIMAS.
- This Project/Thesis or any material, data and information related to it shall not be distributed, published or disclosed to any party by the student except with UNIMAS permission.

(Date) 14 July 2022

Student's signature Supervisor's signature: (12.06.20)

Current Address: Faculty of Resource Science and Technology, University Malaysia Sarawak, Jalan Datuk Mohammad Musa, 94300 Kota Samarahan, Sarawak.

Notes: * If the Project/Thesis is CONFIDENTIAL or RESTRICTED, please attach together asannexure a letter from the organisation with the period and reasons of confidentiality and restriction.

[The instrument was duly prepared by The Centre for Academic Information Services]

Acknowledgements

Motivation and enthusiasm have always been important factors in every venture's success. Many individuals have poured their blessings and heartfelt support on me in favour for this project to be completed successfully. This time, I'm using it to express my gratitude to everyone involved in the project.

First and foremost, I want to convey my heartfelt appreciation to Dr. Lee Kui Soon, my supervisor, and Dr. Samuel Lihan, my co-supervisor. Their passion, patience, insightful remarks, valuable information, practical guidance, and never-ending ideas were invaluable to me throughout my study and writing for my research project. Their vast expertise, extensive experience, and professionalism benefited me in successfully completing this research project. This project would not have been finished without their assistance.

I am also grateful to Encik Abang Iskandharsah bin Abang Hassimsah, the assistant science officer, for his patience and continuous support in obtaining the supplies I required for my project. I also want to express my gratitude to my friends and lab mates for their unwavering support and assistance when I am conducting this project. It would be hard to accomplish this project in such a limited timeframe without their immense support.

Finally, I want to convey my appreciation to my mother, who morally and financially supported me while also offering me advise when I ran into various difficulties during my project. Thank you very much to the postgraduate students in the Virology lab, Sia Siew Chuiang and Khairunnisa Mohammad Hamdi, who assisted me very much in finishing this project with full patience and tolerance. Thank you for all the kind support, patience, guidance and encouragement!

iii

Microbial and Physiochemical Analysis of Water from Semenggok Inland Fishery

Centre, Kuching, Sarawak

Loke Xin Ya

Resource Biotechnology Programme Faculty of Resource Science and Technology University Malaysia Sarawak

ABSTRACT

Water quality is emphasized by the aquaculture sector as it affects the health and growth performance of aquaculture species. However, due to the outbreak of COVID-19 pandemic in Malaysia, various sectors including aquaculture sector, have been instructed to shut down as a lockdown alternative to control the spread of the disease. The total lockdown has made impossible to give proper management to the aquaculture ponds and indirectly led to the high mortality of aquaculture organisms. High mortality has led to rapid microbial growth, which affected the water quality and the balance in the aquaculture system. In this study, the characteristics of aquaculture environmental samples were determined. Water and soil samples were collected from three randomly selected ponds in the Semenggok Inland Fishery Centre, Kuching, Sarawak and the physiochemical and biological parameters were analysed. Bacteria were isolated from both water and soil samples and then characterised. Boiling-centrifugation method was used for DNA extraction of the bacteria. (GTG)₅-PCR was utilized to screen for clonal diversity among the isolates. A dendrogram was constructed using GelJ 1.0 software from the banding profile of (GTG)₅-PCR products. Out of all the isolates analysed, 11 representative isolates were selected for 16S rRNA sequencing based on the grouping from the dendrogram. The 11 isolates were identified as Brevundimonas sp., Staphylococcus sp., Pseudomonas sp., Escherichia sp., Ralstonia sp., and Exiguobacterium sp.. The isolates were tested for antibiotic resistance using the disc diffusion method. Most of the isolates tested were resistant to ampicillin (10 μ g), penicillin (10 μ g), and streptomycin (10 µg). The MAR index of isolates were calculated, ranged from 0.143 to 0.714, indicating high possibility of culturing fish in the contaminated water. This study revealed the risk of presence of multiple antibiotic resistance (MAR) bacteria in the fishery centre. Therefore, the fishery centre should improve the aquaculture system by constantly monitoring and also provide a proper management to the wastewater to minimise the distribution of MAR bacteria.

Keywords: Microbial and physiochemical analysis, (GTG)₅-PCR, 16S rRNA sequencing, Antimicrobial susceptibility testing

ABSTRAK

Kualiti air dititikberatkan oleh sektor akuakultur kerana ia mempengaruhi kesihatan dan prestasi pertumbuhan spesies akuakultur. Berikutan dengan penularan wabak COVID-19 di Malaysia, pelbagai sektor termasuk sektor akuakultur, telah diarahkan tutup bagi mengawal penularan wabak itu. Sekatan ini telah mengakibatkan kekurangan dalam pengurusan kolam akuakultur yang betul dan secara tidak langsung membawa kepada kematian organisma akuakultur. Kematian telah menyebabkan pertumbuhan mikrob yang pesat, menjejaskan kualiti air dan keseimbangan dalam sistem akuakultur. Dalam kajian ini, ciri-ciri sampel akuakultur telah ditentukan. Sampel air dan tanah telah diambil daripada tiga kolam yang dipilih secara rawak di Pusat Perikanan Darat Semenggok, Kuching, Sarawak dan dianalisis parameter fisiokimia dan biologinya. Bakteria telah diasingkan daripada kedua-dua sampel air dan tanah dan bakteria kemudiannya dicirikan. Kaedah pendidihan-sentrifugasi digunakan untuk pengekstrakan DNA bakteria. (GTG)5-PCR telah digunakan untuk menyaring kepelbagaian klon bakteria. Dendrogram telah dibina menggunakan GelJ 1.0 daripada profil banding produk (GTG)₅-PCR. Sebelas pencilan telah dipilih untuk penjujukan rRNA 16S. Daripada 11 pencilan tersebut, terdapat Brevundimonas sp., Staphylococcus sp., Pseudomonas sp., Escherichia sp., Ralstonia sp., dan Exiguobacterium sp.. Pencilan telah diuji untuk rintangan antibiotik menggunakan kaedah resapan cakera. Kebanyakan isolat yang diuji adalah tahan terhadap ampicillin $(10 \mu g)$, penisilin (10 µg), dan streptomycin (10 µg). Indeks MAR bagi pencilan dikira, berjulat antara 0.143 hingga 0.714, menunjukkan kemungkinan tinggi untuk mengkultur ikan dalam air yang tercemar. Kajian ini mendedahkan risiko kehadiran pelbagai bakteria rintangan antibiotik (MAR) di pusat perikanan. Oleh itu, pusat perikanan harus menambah baik sistem akuakultur dengan sentiasa memantau dan juga menyediakan pengurusan yang betul kepada air sisa untuk meminimumkan taburan bakteria MAR.

Kata kunci: Analisis mikrob dan fisiokimia, (GTG)5-PCR, penjujukan rRNA 16S, Ujian kerentanan antimikrob

Table of Content

	Page
Declaration	i
Acknowledgements	
Abstract	iv
Abstrak	iv
Table of Content	V
List of Tables	ix
List of Figures	х
List of Abbreviations	xii
CHAPTER 1: INTRODUCTION	1
CHARPER 2: LITERATURE REVIEW	3
2.1 Water Quality	3
2.1.1 Water Quality in Aquaculture	4
2.2 Physiochemical Analysis	5
2.2.1 Temperature	5
2.2.2 pH	6
2.2.3 Dissolved Oxygen	7
2.2.4 Biochemical Oxygen Demand	8
2.3 Microbial Analysis	8
2.4 Polymerase Chain Reaction	9
2.4.1 Bacterial DNA Fingerprinting Analysis	10
2.4.2 Molecular Identification of Bacteria	10
2.5 Antimicrobial Susceptibility Testing	11
2.6 Challenges of Aquaculture Sectors	12

2.6.1 Aquaculture Diseases Outbreak	12
2.6.2 Global Pandemic	14
2.6.3 Unethical Feeding Practice	15
2.6.4 Media Influences in Aquaculture	15
2.7 Semenggok Inland Fishery Centre	15
2.7.1 Cultured Species	16
2.7.2 Challenges Faced during Pandemic Lockdown	17
CHAPTER 3: MATERIALS AND METHODS	19
3.1 Materials	19
3.2 Methods	21
3.2.1 Study Area	21
3.2.2 Culture Media Preparation	22
3.2.3 Sample Collection	23
3.2.4 Soil Sample Processing	24
3.2.4.1 Preparation of Saline Solution	24
3.2.4.2 Serial Dilution of Soil Samples	24
3.2.5 Microbial Analysis of Samples	25
3.2.5.1 Bacteria Culturing	25
3.2.5.2 Enumeration of Bacteria	26
3.2.5.3 Bacteria Isolation	26
3.2.5.4 Bacteria Stock Preparation	27
3.2.5.5 Bacterial DNA Extraction	29
3.2.5.6 (GTG) ₅ -PCR Fingerprinting	29
3.2.5.7 (GTG) ₅ -PCR Agarose Gel Electrophoresis	30
3.2.5.8 16S rRNA PCR Amplification	30

3.2.5.9 16S rRNA PCR Agarose Gel Electrophoresis	31
3.2.6 Antimicrobial Susceptibility Testing	32
3.2.7 Physiochemical Analysis of Water	
3.2.7.1 Temperature	33
3.2.7.2 pH	33
3.2.7.3 Dissolved Oxygen	33
3.2.7.4 Biochemical Oxygen Demand	33
CHAPTER 4: RESULTS	34
4.1 Microbial Analysis of Samples	34
4.1.1 Enumeration of Bacteria	34
4.1.1.1 Water Samples	34
4.1.1.2 Soil Samples	35
4.1.2 (GTG) ₅ -PCR Fingerprinting	36
4.1.3 16S rRNA PCR Amplification	39
4.2 Antimicrobial Susceptibility Testing	40
4.3 Physiochemical Analysis of Water	43
4.3.1 Temperature	43
4.3.2 pH	43
4.3.3 Dissolved Oxygen	43
4.3.4 Biochemical Oxygen Demand	44
CHAPTER 5: DISCUSSION	45
5.1 Microbial Analysis of Samples	45
5.1.1 Enumeration of Bacteria	45
5.1.2 (GTG) ₅ -PCR Fingerprinting	47
5.1.3 16S rRNA PCR Amplification	50

5.2 Antimicrobial Susceptibility Testing	
5.3 Physiochemical Analysis of Water	55
5.3.1 Temperature	55
5.3.2 pH	55
5.3.3 Dissolved Oxygen	56
5.3.4 Biochemical Oxygen Demand	56
CHAPTER 6: CONCLUSIONS AND RECOMMENDATIONS	57
CHAPTER 7: REFERENCES	58
CHAPTER 8: APPENDICES	62
Appendix 1	62
Appendix 2	64
Appendix 3	66

List of Tables

Page

Table 1:	(GTG) ₅ -PCR fingerprinting reaction mixture for 1 reaction.	29
Table 2:	(GTG) ₅ -PCR fingerprinting protocol.	30
Table 3:	16S PCR reaction mixture for 1 reaction.	31
Table 4:	16S rRNA PCR protocol.	31
Table 5:	Number of total bacteria in water samples.	34
Table 6:	Number of coliform bacteria in water samples.	35
Table 7:	Total number of bacteria in soil samples.	36
Table 8:	The isolation code, bacteria identity and similarity percentage of	40
	the isolate after BLAST analysis.	
Table 9:	Antibiotics resistance and MAR index of bacteria isolates.	42
Table 10:	The temperature of water samples from different sampling ponds.	43
Table 11:	The pH of water samples from different sampling ponds.	43
Table 12:	The amount of DO of water samples from different sampling ponds.	44
Table 13:	The BOD reading of water samples from different sampling ponds.	44

List of Figures

		Page
Figure 1:	Changes in pH during a 24-hour period in waters of an aquaculture	7
	production pond (Fondriest Environmental, 2013).	
Figure 2:	Dissolved oxygen tolerance range for fish (Leaffin, 2019).	8
Figure 3:	Approximately 1.5 kb 16S rRNA gene of E.coli showing the nine	9
	variable regions that make it an ideal target as a phylogenetic	
	marker gene (Ashram et al., 2017).	
Figure 4:	Tilapia Fish (Oreochromis Niloticus) (Tolentino et al., 2020).	16
Figure 5:	Lampan Jawa (Barbonymus gonionotus) (Amang, 2021).	16
Figure 6:	Tengadak (Barbonymus schwanenfeldii) (Amang, 2021).	17
Figure 7:	Geographic coordinates of Semenggok Inland Fishery Centre and	21
	the site of water and soil sampling. Source: Google Maps.	
Figure 8:	First pond selected for sample collection, labelled as Pond 1 (P1).	21
Figure 9:	Second pond selected for sample collection, labelled as Pond 2 (P2).	22
Figure 10:	Third pond selected for sample collection, labelled as Pond 3 (P3).	22
Figure 11:	The labelled water samples collected from the fishery centre.	23
Figure 12:	The labelled soil samples collected from fishery centre.	24
Figure 13:	Bacteria culturing onto NA and EMB agar via spread plate method	25
	aseptically for water samples.	
Figure 14:	Bacteria culturing onto MHA agar via spread plate method	26
	aseptically for soil samples.	
Figure 15:	Streak plate of water samples.	27
Figure 16:	Streak plate of soil samples.	27
Figure 17:	Bacteria stock in MH slant agar for water samples.	28

Figure 18:	Bacteria stock in MH slant agar for soil samples.	28
Figure 19:	Bacteria stock in LB broth for water samples.	28
Figure 20:	Bacteria stock in LB broth for soil samples.	28
Figure 21:	First run of agarose gel electrophoresis for (GTG) ₅ -PCR	36
	fingerprinting product.	
Figure 22:	Second run of agarose gel electrophoresis for (GTG) ₅ -PCR	26
	fingerprinting product.	
Figure 23:	Third run of agarose gel electrophoresis for (GTG) ₅ -PCR	37
	fingerprinting product.	
Figure 24:	Fourth run of agarose gel electrophoresis for (GTG) ₅ -PCR	37
	fingerprinting product for samples does not show clear band during	
	third run.	
Figure 25:	A dendrogram illustrating the relationship between the bacterial	38
	isolates analysed by repetitive sequence based PCR using (GTG) ₅	
	primer.	
Figure 26:	Results of 1.5% (w/v) agarose gel electrophoresis of 16S rRNA	39
	PCR product.	

- Figure 27: Results of 1.5% (w/v) agarose gel electrophoresis of 16S rRNA 39 PCR product for sample 21 and sample 29.
- Figure 28:The positive control of antimicrobial susceptibility testing by using41*Escherichia coli* ATCC 25922 strains.
- Figure 29: Antibiotic susceptibility testing of isolate 20, *Ralstonia* 41 *mannitolilytica*.

List of Abbreviations

°C	degree Celsius
Amp	Ampicillin
AST	Antimicrobial Susceptibility Testing
ATCC	American Type Culture Collection
BLAST	Basic Local Alignment Search Tool
BOD	Biochemical Oxygen Demand
bp	Base Pair
С	Chloramphenicol
CFU	Colony-Forming Unit
Cl ₂	Chlorine
COD	Chemical Oxygen Demand
COVID-19	Coronavirus Disease 2019
CO_2	Carbon Dioxide
dH ₂ O	Distilled Water
DNA	Deoxyribonucleic Acid
dNTP	Deoxynucleotide Triphosphate
DO	Dissolved Oxygen
E	Erythromycin
EDTA	Ethylenediaminetetraacetic Acid
EMBA	Eosin Methylene Blue Agar
EtBr	Ethidium Bromide
Κ	Kanamycin
kb	Kilobase
LB	Luria-Bertani

MAR	Multiple Antibiotic Resistance
MgCl ₂	Magnesium Chloride
MHA	Muller Hinton Agar
MIC	Minimum Inhibitory Concentration
Na	Nalidixic Acid
NA	Nutrient Agar
NC	Negative Control
NCBI	National Center for Biotechnology Information
NH ₃	Ammonia
O ₂	Oxygen
Р	Penicillin
PCR	Polymerase Chain Reaction
pH	Potential of Hydrogen
ppm	parts per million
RNA	Ribonucleic Acid
rRNA	Ribosomal Ribonucleic Acid
rRNA S	Ribosomal Ribonucleic Acid Streptomycin
S	Streptomycin

CHAPTER 1

INTRODUCTION

The aquaculture industry has constantly received the attention of the Malaysian government since it has been highlighted as a priority area in the country's economic growth (Jumatli & Ismail, 2018). Furthermore, the government and the public have always emphasized water quality in aquaculture as it is a foundation of health and well-being. Water quality is one of the most critical elements affecting the health and performance of aquaculture farms (Tower, 2015). The water quality can influence the health and growth of aquaculture species, but it also has the potential to affect people, making it zoonotic.

Water quality is described as a water body's chemical, physical, and biological qualities, typically linked to its long-term viability (Roy, 2018). It is critical in aiding scientists in predicting and determining the impact on humans and the environment. It is also significant as an assessment effort in restoration projects to maintain environmental standards. As the general population is aware, water is vital for the survival of living organisms and many activities. The decent quality of water will be free of contaminants and safe to drink for living organisms. On the other hand, poor water quality has the most significant direct influence on aquatic animals, notably fish, which is linked to food supplies.

We shall not be able to identify the quality of the water merely by looking at it with our bare eyes. To assess the water quality, specific procedures and analysis are necessary. Scientists will be able to determine the water quality using a variety of indicators. The physiochemical examination of water can provide these indicators. Not only that, but microbes are another critical factor that influences water quality. Microorganisms in the water body will enable the analysis of microbial part. Although the microorganisms in the water may not be hazardous, there is a possibility that they are pathogenic and can cause harm to humankind. Back to the end of 2019, due to the outbreak of the COVID-19 pandemic, various sectors in Malaysia, including the aquaculture sector, have been instructed to shut down as a lockdown alternative to control the spread of the disease. The total lockdown has resulted in a shortage of manpower to give proper management to the aquaculture ponds as people were restricted from any outdoor activities. This indirectly led to the high mortality of aquaculture organisms. The dead aquaculture carcasses left in long-term unmanaged aquaculture ponds provided an opportunity for rapid microbial growth, which further affected the water quality and the balance physiochemical parameter of the aquaculture pond.

Therefore, in this research project, the microbial and physiochemical study of water and soil samples obtained from Semenggok Inland Fishery Centre was conducted. The physiochemical parameters of the water samples were examined. Following the water assessments, the collected data was compared to the Malaysia's National Water Quality Standard. Microbial analysis was performed on the water and soil samples collected to identify their microbial composition. Laboratory techniques such as (GTG)₅-PCR fingerprinting and agarose gel electrophoresis (AGE) were employed to aid in the investigation of the microbial content of the samples to detect their microbial profile. The bacteria species found in the water and soil samples were identified using 16S rRNA sequencing. Finally, antimicrobial susceptibility test (AST) was performed to assess the antibacterial characteristics of different isolates from water and soil samples against commonly used antibiotics in the fish farming industry. It is hypothesised that the water quality from the fishery centre is suitable to support the survival of aquatic life.

The objective of this project is to examine the water quality of selected ponds at Semenggok Inland Fishery Centre via physiochemical and microbial analysis. This project also aims to determine and identify the bacteria population in examined water samples. Lastly, this project seeks to determine the antimicrobial susceptibility patterns of identified isolates.

CHAPTER 2

LITERATURE REVIEW

2.1 Water Quality

Water quality is described as the chemical, physical, and biological qualities of water, typically linked to its long-term viability (Roy, 2018). It is crucial for monitoring and analysing the impact on human being and the environment. There are three types of metrics used to evaluate water quality: physical, chemical, and biological (Omer, 2019).

Physical water body measurements included assessing the appearance of water bodies, which often included colour, turbidity, taste, odour, and temperature (Omer, 2019). The water body must be free of any contaminants that are objectionable to the senses of sight, taste, or smell to be used for a specific purpose, and the most essential physical attribute that should be addressed while assessing water quality is turbidity (Abu Shmeis, 2018). It included processes for measuring parameters such as flow conditions, substrates, and pollutants that have a direct impact on the quality of aquatic environments.

Chemical measurements are concerned with the measurement of controlled chemical compounds or combinations of chemicals that have the potential to occur in a water body at levels that are detrimental to living organisms. To assess water quality, hazardous contaminants, pH, dissolved oxygen (DO), biochemical oxygen demand (BOD), chemical oxygen demand (COD), suspended solids, nutrients, metals, synthetic organic compounds, and radionuclides were all often collected or monitored (Omer, 2019).

The assessment of a water body's biology for its potential to give biological protection to living organisms is referred to as biological measurements of water. It is a method of determining how healthy a body of water is. The evaluation of the integrity of an aquatic ecological system that results from a biological inventory and biological potential analysis is commonly termed a biological assessment of water quality. The presence of microorganisms such as bacteria, algae, protozoa and viruses are the parameters for biological measurements (Omer, 2019).

Aside from physiological, chemical, and biological aspects, climate, vegetation, geology, and human activities all have an impact on water quality. These are known as natural and anthropogenic factors (Akhtar et al., 2021). The quality of water is frequently affected by these human and natural effects. Human activities such as mining, for example, are point sources of pollution that endanger water quality. Non-point sources, such as pollution, will also harm the water in the long and short-term owing to the pollutants that are released into the ecosystem and generate undesired impacts or squander resources. Water quality is also affected by faeces-related pollutants, agricultural pollutants, air pollutants, pathogens, and chemical contaminants.

2.1.1 Water Quality in Aquaculture

When it comes to growing the aquatic organisms, water quality is crucial. The appropriate water quality varies by species and must be evaluated to ensure the development and survival of aquatic organisms. The quality of the water used has an impact on the health and performance of aquatic organisms, as well as the cost of them in the market.

The water quality in the aquaculture industry is constantly examined by government. Temperature, dissolved oxygen (DO), pH, biochemical oxygen demand (BOD), ammonia (NH₃), and nitrates are all regularly measured parameters. Carbon dioxide (CO₂), chlorine (Cl₂), and salinity are occasionally monitored as well, depending on the culture system.

In Malaysia, the government established a set of water quality regulations based on the class of water and its intended use. The regulation is called "National Water Quality Standards for Malaysia" (*Appendix 1*). According to the standards, aquaculture sector can be categorized as class III water, with the uses stated as for common and economic value and tolerant species, livestock, and drinking. The parameter measured for class III water is NH₃, BOD, COD, DO, pH, total suspended solid, temperature, faecal coliform, and the total coliform in the water.

2.2 Physiochemical Analysis

The examination of the water's physical and chemical characteristics is referred as the physiochemical analysis of the water. It is important to create an appropriate living environment for aquatic organisms in ensuring their growth quality and continuous production. There are various physiochemical parameters, such as pH, temperature, dissolved oxygen (DO), and biological oxygen demand (BOD).

2.2.1 Temperature

The temperature of a water system is crucial because it affects the development and reproduction of aquatic species and the rate of all chemical reactions in a well-established water system (Patil et al., 2012). Temperature fluctuations in aquaculture system can harm aquatic species by affecting their metabolism, the degree of ammonia poisoning, and feeding rates. The rate of metabolism, biological and chemical reactions, and oxygen consumption will double for every 10°C increase in temperature. Temperature conditions, however, differ depending on the species. The temperature substantially impacts the biota respiration, such as oxygen (O₂) consumption rates. This is because warmer water contains less O₂ than cooler water.

Fishes are particularly vulnerable to rapid temperature fluctuations since the fishes can modify their body temperature to suit the environment. The water temperature can affect every element of their survival, including breathing and reproduction. Water enters the fish through the mouth and is driven out through the gills during breathing. The DO in the water will enter the blood cells of the fish. The amount of O_2 in the water decreases as the temperature rises, and the amount of O_2 given to the fish decreases as well. As a result, warmer water is incompatible with fish survival. The fish cannot breathe if the temperature rises to the dangerous levels.

Water temperature also significantly impact the reproduction system of fish since different kinds of fish have different needs for reproduction. For example, salmon and other cold-water fish spawn at low temperatures, whereas warm-water fish reproduce at a higher temperature. By these preferences, an aquatic habitat that sees a significant temperature shift that is out of character for it might lead fish to either depart or decrease in number owing to a lack of reproduction.

2.2.2 pH

The term pH is derived from a French phrase "*pouvoir hydrogéne*", indicating the meaning of "hydrogen power". This parameter is critical in determining whether water is corrosive as a living environment of aquatic organisms (Patil et al., 2012). Maintaining a constant pH within a healthy range is crucial since it affects the metabolism and other physiological activities of aquatic organisms. The ideal pH levels for fish farming systems should vary between 6.5 to 9.0 (Fondriest Environmental, 2013). Beyond the ideal range, stress, illness susceptibility, decreased productivity, poor growth, and even mortality may occur. Fishes are sensitive to pH shifts in the high temperature reading water body, mainly due to the amount of CO₂ produced in the water during the daytime that impacts the pH. The pH of the water will naturally change during the daytime as the phytoplankton will use CO₂ for photosynthesis. Affecting by the amount of sunlight during daytime, the pH value of the water is the lowest before sunrise and highest in the afternoon (*Figure 1*).

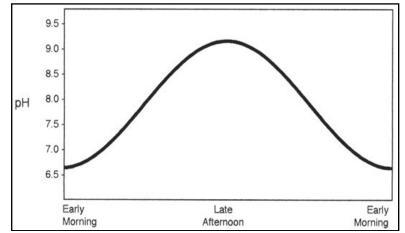


Figure 1. The pH changes in an aquaculture producing pond's waters throughout a 24-hour period (Fondriest Environmental, 2013).

2.2.3 Dissolved Oxygen

Measurement of dissolved oxygen (DO) is also significant in aquaculture sector. It is the amount of oxygen that can be accessible by all the aquatic organisms in the water. Optimal DO level is crucial for optimum aquatic production since it directly influences their feed intake, illness resistance, and metabolism. When DO is scarce in the water, the aquatic species will become incredibly stressed. A DO level lower than 3 parts per millions (ppm or mg/L) will start to stress the growth and weaken the immune response of fish species; and levels below 1 mg/L will be lethal. A suitable DO levels for fishery aquaculture is suggested at above 4 mg/L (*Figure 2*).

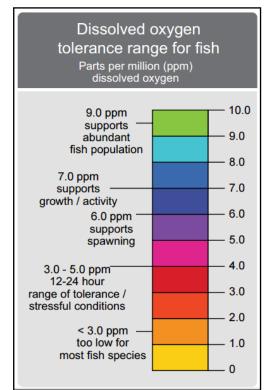


Figure 2. Dissolved oxygen tolerance range for fish (Leaffin, 2019).

2.2.4 Biochemical Oxygen Demand

The aquaculture pond oxygen balance can also be influenced by biochemical oxygen demand (BOD). It is described as the amount of DO in a constant temperature that the aerobic organisms in the water system ought for organic substances break down. BOD is usually measured using the five-day biochemical oxygen demand (BOD₅). It is a significant water quality indicator that is often required to obtain with government-issued water quality permits and farm certification. The quantity of DO deplete quickly when the BOD value was high, which will affect the fish's development and production.

2.3 Microbial Analysis

The microbial analysis is also vital in the aquaculture farming system to ensure the survival of aquatic organisms. In an aquaculture system, microbes such as *Staphylococcus* sp., *Bacillus* sp., *Streptococcus* sp., *Pseudomonas* sp., *Escherichia* sp., and *Citrobacter* sp. are often detected (Ajayi & Okoh, 2014). Microbes have beneficial role in pond system maintenance, notably in production, nutrient cycling, cultured animal feeding, water quality,

and environmental run-off effect. As for pathogenic microorganisms, they may undermine the fish development, causing a myriad of problems to the industry (Nova Biologicals Teams, 2018; Sandle, 2016). As a result, microbial analysis is crucial for the early detection of the aquaculture water environment before an irreversible loss occurs.

Escherichia sp., *Enterobacter* sp., and *Citrobacter* sp. are the example of coliform bacteria, or known as Enterobacteriaceae. They are rod-shaped, gram-negative, and does not produce spores. Coliform bacteria do not cause disease. However, when assessing the water quality, the coliform bacteria can act as an excellent indicator of the existence of infective microbes. Excessive levels of faecal coliform indicate the failure in performing water treatment. Therefore, antibiotics are used in aquaculture sector in preventing the excessive growth of pathogenic microbes. Antimicrobial susceptibility testing (AST) is important to identify the inhibition effects of the growth of microorganisms that causing the infection to the aquatic species by using different types of antimicrobials on Muller Hinton (MH) agar.

2.4 Polymerase Chain Reaction

Polymerase chain reaction (PCR) is a technique that often applied in the field of biotechnology. This technique will amplify the DNA segment for various laboratory applications. In the early 1980s, Mullis and associates created PCR, with the basis of Panet and Khorana's successful *in-vitro* amplification of DNA (Ghannam & Varacallo, 2022). This technique were later be awarded with a Nobel prize just a decade later. As this technique is useful due to the ability to amplify the specific regions of DNA into billion folds, it is used in many applications, such as gene cloning, infectious diseases diagnosis and genetic abnormalities screening (Ghannam & Varacallo, 2022).

To perform PCR, there are few main components required, which is the PCR primers, DNA polymerase, DNA templates and free nucleotide bases. The DNA template contains a specific region of DNA that wanted to be amplified. As for primers, it is oligonucleotides