Synthesis of Silver (I) Coordination of Aspirinate Azo Ligands as Potential Antibacterial Agents

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Abstract. The rise of antimicrobial resistance for infectious bacteria has become an alarming issue to human health. New antimicrobial drugs are in dire need and pivotal to overcome this issue. In this study, aspirinate azo ligands bearing different halogens L1-5 has been prepared *via* diazo-coupling reaction. The ligands L1-5 were coordinated with silver, Ag (I) metal to produce Ag(I) aspirin-azo complexes C1-5. The antibacterial properties of L1-5 and C1-5 were evaluated against *Staphylococcus aureus* and *Escherichia coli* using turbidimetric kinetic method. The complexes C1-5 showed comparable growth inhibition activity towards *E. coli* (MIC 82-105 ppm) and *S. aureus* (MIC 80-105 ppm) compared to ligands L1-5 with *E. coli* (MIC 83-200 ppm), *S. aureus* (80-131 ppm) and ampicillin (MIC 93 and 124 ppm, respectively). The excellent bacterial resistance of both L1-5 and C1-5 indicates the potential of aspirinate azo and their complexes as new antibacterial agents, which significantly benefit to the pharmaceutical industries.

Introduction

The increase of contagious bacteria has led to the emergence of drug resistance which caused many deaths in the society [1]. The development of new drugs with antimicrobial drug resistance properties is in dire need to overcome the growth of pathogenic bacteria. Several types of antimicrobial drugs have been developed through the manufacture of commercial drugs derived from azo dyes such as phenazopyridine [2].

Several studies have been reported on the synthesis of azo dyes with a wide spectrum of biological activity [3]. The azo moieties have the ability to prevent bacteria multiplication and DNA function [4, 5]. Organic dyes featuring azo moieties (N=N) in the molecular network have also been widely studied due to their thermal and energy storage properties [6, 7, 8].

Hybridization of azo dyes with natural product-based compounds with multifunctional groups can increase the efficiency and probability of additional interactions with biological targets/receptors [3]. Our preliminary studies on azo dyes bearing natural product-based compounds such as aspirin and kojic acid showed antibacterial activities [3, 8, 9, 10] and anticancer activity [11, 12].

Complexation of azo dyes with heavy metals namely Ag(I) can also increase the cytotoxic and antibacterial properties [13]. The formation of chelation between a metal ion and ligand able to improve the biological activities compared to the ligands alone [14]. Complexation is able to increase the lipophilicity of ligands, which allow the ligand to enter the bacterial cell and interact with the enzyme receptor or DNA [13]. Ag metal is well known for its antimicrobial properties [15, 16,17]. Ag is non-toxic to humans and has been utilized as antimicrobial agents due to its ability to interact with protein and enzyme of bacteria, which eventually leads to bacterial cell death [18, 19, 20].

In this paper, the synthesis of natural product-based compounds namely aspirinate azo ligands L1-5 bearing halogens (F, Cl, Br, I) were synthesized *via* diazotization and coupling reaction. The complexes C1-5 were prepared *via* complexation with Ag(I) in dark condition and at room temperature. The ability of the ligands and complexes to inhibit the growth of *S. aureus* and *E. coli* were tested using turbidimetric kinetic method.

Experimental

Materials and instrumentational

Chemicals and solvents were used as received from Merck. The analysis of melting point was performed using open capillary method on Stuart SMP3. Smart Omni Transmission Nicolet IS10 Perkin Elmers Thermoscientific was used to analyze FTIR spectra. ¹H and ¹³C NMR spectra were obtained from JEOL ECA with ¹H at 500 MHz and ¹³C at 125 MHz using standard reference, DMSOd₆. The elemental CHNS was analyzed using Thermo-Flash (EA 1112) CHN Analyser. The transmittance values were analyzed on Optima SP-300.

Synthesis of halogenated aspirin-azo ligands (L1-5)

Sodium nitrite (1M, 2mL) was poured into a solution of aniline derivatives (5 mmol) and hydrochloric acid (8 M, 8 mL) in an ice bath. Acetylsalicylic acid (5 mmol) and sodium hydroxide (1 M, 10.0 mL) were added and continue stirring for 45 min. Hydrochloric acid (3 mL, 8 M) was added dropwise onto the solution mixture until the precipitate formed. The solid was filtered, rinsed, recrystallized from hot ethanol and dried under vacuum to obtain the final product L1-5.

Complexation of Ag (I) complexes [Ag(Aspirin-azo)]NO₃ (C1-5)

Silver nitrate (1 mmol) was added dropwise into aspirin-azo ligands L1-5 (0.5 mmol) in methanol. The mixture of silver nitrate and ligand was stirred for 3 h in the dark at room temperature. Solid precipitates formed were filtered and washed with water followed methanol and dried to afford the final product C1-5.

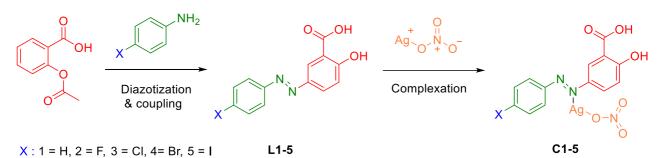
Antibacterial study

Antibacterial studies of the synthesized aspirinate azo ligand L1-5 and Ag(I) complexes C1-5 were carried out against *E. coli* (ATCC 25922) and other common types of bacteria, *S. aureus* (S48/81). Both bacteria sub-cultures were grown in Luria-Bertani (LB) broth at 37 °C.10 mL of LB broth was added with ligands and complexes at different concentrations separately (50, 80 and 100 ppm). The media and tested compounds were added with 0.2 mL of sub-cultured bacteria. The mixture was then stirred and incubated at 37 °C. Dimethyl sulfoxide and ampicillin were set as controls, negative and positive, respectively. The transmittance (T) values were recorded for every 1 h (0h – 6h) at wavelength 560 nm. The T values were extrapolated to colony-forming units / mL and articulated in ln Nt².

Results and Discussion

Synthesis

Aspirinate azo ligands L1-5 were initially synthesized *via* diazotization of halogenated anilines derivatives followed by coupling with acetylsalicylic acid with yields 24-85 %. Ag(I) complexes C1-5 were prepared satisfactorily from direct reaction of L1-5 with silver nitrate in a molar ratio of 1:2 (Ag: ligand) to produce 26-68 % yields, where azo was acted as nitrogen chelator¹. The equivalent ratio of Ag(I) and azo ligand gave similar composition and equivalent yields. The synthetic pathway of aspirinate azo ligands L1-5 and Ag(I) complexes C1-5 is shown in Scheme 1. The structural conformation of ligands L1-5 and Ag(I) complexes C1-5 was performed using CHN, FTIR and NMR spectroscopy.



Scheme 1. The synthetic pathway of halogenated azo-aspirin L1-5 and Ag (I) azo-aspirin complex C1-5.

Based on CHN analysis, the data obtained for L1-5 and C1-5 corresponded to the proposed structure of the ligands and complex, [Ag (Aspirin-azo)]NO₃. The key functional group present in the IR spectra L1-5 and C1-5 is depicted in Table 1. The IR spectra of L1-5 showed a broad band in the region 1437–1487 cm⁻¹, which attributed to the v(N=N) [3]. The peaks presence at 1619-1688 cm⁻¹ and 1660–1676 cm⁻¹ were attributed to v(C=O) vibration, while the appearance of peaks at 1561–1591 cm⁻¹ was attributed to v(Ar) stretching. Upon coordination of azo ligands with silver(I) ions, the FTIR peaks of C1-5 were significantly shifted indicates successful coordination to produce new silver(I) aspirinate azo compounds. All the absorption peaks attributed to v(C=O), v(Ar) and v(N=N) in C1-5 were shifted to a higher frequency [21, 22].

The significance resonance of ¹H and ¹³C NMR corresponded to the L1-5 and C1-5 structure is shown in **Table 2** and **Table 3**. The coordination of Ag(I) complex C1-5 is shown in Fig. 1.

The coordination of Ag(I) with aspirinate azo ligands L1-5 was confirmed by the shifted of peaks in the spectra. The peaks corresponded to H₆ in L1-5 were assigned as a doublet at δ 8.30 -8.33 ppm and H₄ as doublet to doublet at δ 7.92-8.07 ppm, which strongly indicated on the successful coupling reaction *via* the formation of N=N at the *para* position of OH group. Upon coordination with Ag(I) ions, H₄ was shifted to the upfield at δ 0.06-0.21 ppm. While H₂, was shifted upfield at δ 0.01-0.10 ppm. The shift of ¹H peaks was also contributed by the existence of nitrate ions (NO₃⁻) in the complexes. The nitrate ions worked as a counter ion and established a small or minor interaction with an aromatic proton [23].

The ¹³C NMR spectra of **L1-5** and **C1-5** were in agreement with the target structures as indicated by the occurrence of resonances at δ 148.5-152.2 ppm and δ 142.7-144.4 ppm attributed to C₁, and C₅, respectively. Upon coordination with Ag(I) ions, C₅ has shifted upfield in the range of δ 0.82-1.58 ppm. While C₁, was shifted downfield in the range of δ 0.21-0.49 ppm (Table 3). Ag(I) was envisaged to coordinate with N bonded to C₅ due to the strong shifting effect [24, 25].

Compound	Wavelength (cm^{-1})		
	v(C=O)	v(Ar)	v(N=N)
L1	1688	1587	1480
C1	1661	1561	1450
L2	1666	1591	1487
C2	1624	1561	1444
L3	1659	1576	1478
C3	1661	1580	1458
L4	1665	1574	1455
C4	1619	1561	1437
L5	1661	1584	1465
C5	1622	1563	1440

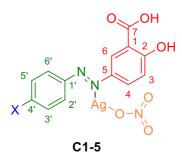
Table 1. Summary of main FTIR band of azo ligand L1-5 and Ag(I) complex C1-5.

Compound	¹ H NMR shifts in ppm		
Compound			
	H4	H ₆	H ₂ ,
L1	7.920	8.306	7.840
C1	7.857	8.330	7.821
Shift value (ppm)	0.063	0.024	0.019
L2	8.071	8.321	7.947
C2	7.862	8.326	7.896
Shift value (ppm)	0.209	0.005	0.051
L3	8.077	8.332	7.880
C3	7.991	8.318	7.870
Shift value (ppm)	0.086	0.014	0.010
L4	8.082	8.330	7.808
C4	7.881	8.316	7.772
Shift value (ppm)	0.201	0.014	0.036
L5	8.077	8.234	7.951
C5	7.915	8.310	7.848
Shift value (ppm)	0.162	0.080	0.103

 Table 2. Summary of ¹H NMR resonances nearest to the N=N moieties.

Table 3. The resonances	of 13 C NMR for L1-5 and C1-5.
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Compound	¹³ C NMR peak shifts in ppm		
	C_1 '-N	C ₅ -N	
L1	151.814	144.460	
C1	152.205	142.895	
Shift value (ppm)	0.391	1.385	
L2	148.580	144.288	
C2	148.914	142.924	
Shift value (ppm)	0.334	1.364	
L3	150.412	144.298	
C3	150.621	143.477	
Shift value (ppm)	0.209	0.821	
L4	150.707	144.326	
C4	151.117	142.829	
Shift value (ppm)	0.410	1.497	
L5	151.117	144.298	
C5	151.604	142.714	
Shift value (ppm)	0.487	1.584	



X : 1 = H, 2 = F, 3 = Cl, 4= Br, 5 = I **Figure 1**. Ag(I) azo-aspirin complex **C1-5** structure.

Antibacterial analysis

Aspirinate azo ligands L1-5 and Ag(I) aspirinate azo complexes C1-5 showed antimicrobial properties screened on bacteria *S. aureus* and *E. coli* at the various concentration [26]. In both bacteria, MIC values were obtained by inducing the growth rate concentration to zero for both bacteria as shown in **Table 4**.

Ligand/complex	E. coli	S. aureus
L1	117	104
C1	82	104
L2	>200	125
C2	105	131
L3	85	98
C3	91	80
L4	86	70
C4	91	81
L5	83	80
C5	90	72
Ampicillin	93	124

Table 4. MIC values of L1-5 and C1-5 for E. coli and S. aureus.

Ligands L1, L3-L5 demonstrated good antibacterial activities with MIC values 83-117 ppm and 70-104 ppm against *E. coli* and *S. aureus* ppm respectively, which are comparable to ampicillin (MIC 93 ppm and 124 ppm, respectively). Aspirinate azo dyes L5 bearing iodine exhibited excellent inhibition against *E. coli* with MIC 83 ppm, while L4 bearing bromine showed excellent growth inhibition against *S. aureus* with MIC 70 ppm. The presence of azo moieties in the ligands has contributed to the strong inhibition of bacteria growth [27, 28]. Additionally, the presence of hydroxyl (OH) and carboxyl group (COOH) in the molecular network has contributed to the hydrophilic properties of the molecules which allowed the ligand to interact with bacteria receptors or DNA *via* hydrogen bonds interaction. The halogen in the ligand has contributed to the increase of antimicrobial activity. Moreover, the halogen has the capability to upsurge the lipophilicity of ligand for facile diffusion into the bacterial cell wall and membrane *via* the interaction of halogen bond towards bacterial receptor or DNA [29, 30, 31].

The formation of Ag(I) complexes, however, showed the increase and decrease in the antimicrobial activities (Table 4). The complexes (C1-5) gave comparable and lower MIC values of 82-105 ppm and 72-131 ppm against *E. coli* and *S. aureus*, respectively compared to ampicillin (MIC 93 ppm and 124 ppm, respectively) and the ligands L1-5. While the complexes (C1-2) demonstrated a stronger ability to inhibit the growth of *E. coli* with MIC 82 ppm and 105 ppm, respectively compared to their free ligands L1 (117 ppm) and L2 (>200 ppm). While the growth inhibition activity against *S. aureus* demonstrated compound C3 and C5 with excellent MIC values of 80ppm and 72 ppm, respectively, in comparison to their ligands L3 (MIC 98 ppm) and L5 (MIC 80 ppm).

Upon coordination with Ag, the antibacterial activity was increased as the lipophilicity of the compound increased [31]. Furthermore, Ag(I) ions have also improved the π -electrons delocalization throughout the ligand molecule, thus increase its ability to penetrate the lipid cell membrane of the bacteria [32, 33]. The decrease in the inhibition activities is believed due to the high lipophilic properties of the complexes after coordinated with Ag that reduced the solubility of complexes to dissolve and react with enzyme and protein [34]. It can be concluded that the combined interaction of hydrophilicity and lipophilicity of a molecule plays an essential part in the antibacterial properties of the compounds [3, 32].

Conclusion

Aspirinate azo derivatives bearing series of halogens L1-5 were successfully synthesized and coordinated with Ag to form complexes C1-5. The azo derivatives L4 gave excellent inhibition against *S. aureus* (MIC 70 ppm) while complex C1 showed excellent inhibition against *E. coli* featuring the lowest MIC value (MIC 82 ppm) as compared to other ligands, complexes and standard ampicillin antibiotic drug. The presence of halogens in L2-5 exhibited excellent antimicrobial activities compared to L1. Coordination of ligands with Ag(I) has increased the lipophilicity of the overall complexes and increased the antibacterial activity. C3-5 had a marginal decrease in activity for growth inhibition against *E. coli* as compared to the ligands. The excellent antibacterial properties of aspirinate azo dyes and Ag coordination have shown promising potential for the development of new antibiotic drugs in the pharmaceutical industry.

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