

## Nomenclatural Notes on a Caecilian (Amphibia: Gymnophiona) Name

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Bhatta and Srinivasa (2004) described a new species of caecilian, which they named *Gegeneophis madhavai*, from “Doddingaguli locality (Mudur Village, Kundapura Taluk, Udipi District, Karnataka State).... “situated adjacent to the Mookambika Wildlife Sanctuary in the Western Ghats” (in southwestern India).

The new species was named for four individuals who share a first name, namely, Madhava Bhat, Madhavarao Bhide, Madhava Anantha and Madhava Gadgil. The spelling of the first (= given) name of the last individual is, in fact, Madhav, a contemporary north Indian version of the spelling. The name is derived from the Sanskrit, meaning vernal, originating from the name Madhu, a legendary king, whose most famous descendant is Krishna, a Hindu god (see Hanks and Hodges 1990).

According to Article 32.5 of the International Code of Zoological Nomenclature (International Commission of Zoological Nomenclature 1999), in case of clear inadvertent error in the formation of a new nomen, correction of the nomen is possible (see Bauer and Das 2000 and Michels and Bauer 2004, for some recent examples of corrections of original nomen). Indeed, Article 34.2 states that errors in spelling of the endings must be corrected. In the present case, because the new caecilian is named for more than a single individual, the termination of the species nomen should be *-orum*. The species nomen of the south Indian caecilian described by Bhatta and Srinivasa (2004) is here emended to *Gegeneophis madhavaorum*.

Under Article 33.2.2, the corrected nomen retains the original authorship and date. The valid name of the new south Indian caecilian is therefore *Gegeneophis madhavaorum* Bhatta and Srinivasa, 2004.

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## A Male Hybrid from *Aspidoscelis sonorae* (Parthenogenetic) and *A. burti stictogramma* (Bisexual)

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Although *Aspidoscelis burti stictogramma* (Giant Spotted Whiptail) is remarkable for its past participation in hybridizations contributing to the origins of six triploid parthenogenetic species: *A. exsanguis*, *A. flagellicauda*, *A. opatae*, *A. sonorae*, *A. uniparens*, and *A. velox* (Reeder et al. 2002; Wright 1993), contemporary hybridization by *A. burti stictogramma* is undocumented. Participation in hybridization events by the six parthenogenetic species ranges from rare to absent, with the frequency determined by opportunity and ecological context (Taylor et al. 2001; Walker et al. 1989). Rare hybridization is exemplified by *A. exsanguis*, *A. sonorae*, *A. uniparens*, and *A. velox* (Cuellar and McKinney 1976; Hardy and Cole 1998; Neaves 1971; Taylor et al. 1989), and hybridization by *A. flagellicauda* and *A. opatae* is unknown—because contact with bisexual congeners is limited for the former (Wright and Lowe 1968) and absent for the latter (Wright 1967). The objective of this report is to describe the first known hybrid between *A. sonorae* and *A. burti stictogramma*.

On 12 June 1972, P. Casella collected a female *A. burti stictogramma* (119 mm snout–vent length [SVL]; Regis University [RU] 7232) and four specimens that I identified as parthenogenetic *A. sonorae* (RU 7233–7236). These five specimens were collected in a lush, Lower Sonoran Zone habitat (Lowe 1964, his Fig. 6) on a lower slope of the Santa Catalina Mountains (accessed from the Mount Lemmon Highway) east of Tucson, Arizona. In 1992, I dissected each specimen for evidence of egg clutches and was surprised to find normal appearing testes and vasa deferentia in RU 7234, a specimen of 70 mm SVL (Fig. 1). Its testes dimensions (left: 5.2 × 3.0 mm, right: 5.5 × 3.2 mm) were comparable in relative size to the testes (left: 5.4 × 4.4 mm, right: 6.5 × 4.0) of an adult male *A. burti stictogramma* of 87 mm SVL (RU 95170, Fig. 1) collected on 9 July 1995 from a different locality. The ratio of testis width to SVL for RU 95170 was within the 95% confidence interval expected for males of *A. burti stictogramma* collected in July (Goldberg 1987). Although RU 95170 (87 mm SVL) was likely reproductively mature, it still expressed the juvenile color pattern of *A. burti stictogramma*—pale gray, intact stripes sharply contrasting with the adjacent dark brown fields, each field supporting small tan spots, and a subdued dorsal pattern on the legs (Fig. 1). In contrast, the representative of *A. burti stictogramma* collected at the hybridization site (RU 7232, 119 mm SVL) had completed the ontogenetic transition—fragmentation of primary stripes into large spots, enlargement and lightening of dark field spots, and establishment of a high density of prominent pale spots on the dorsal surfaces of the legs—to a final color pattern characterized by spots rather than by stripes.

There were two reasons why RU 7234 could not be reassigned to *A. burti stictogramma*. First, the male had only 70 granular scales