

Characterization of coenzyme binding and selectivity determinants in *Mycobacterium tuberculosis* flavoprotein reductase A: analysis of Arg199 and Arg200 mutants at the NADP(H) 2_-phosphate binding site

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Mycobacterium tuberculosis FprA (flavoprotein reductase A) is an NAD(P)H- and FAD-binding reductase that is structurally/ evolutionarily related to adrenodoxin reductase. Structural analysis implicates Arg₁₉₉ and Arg₂₀₀ in interactions with the NADP(H) 2_-phosphate group. R199A, R200A and R199A/ R200A mutants were characterized to explore the roles of these basic residues. All mutations abolished neutral FAD semiquinone stabilization in the NADPH-reduced enzyme, owing to weakened NADPH affinity. Instead, FAD hydroquinone was formed in all mutants, and each displayed substantially enhanced autooxidation rates (20– 40-fold) compared with NADPH-reduced WT (wild-type) FprA. Steady-state ferricyanide reduction studies revealed diminished NADPH affinity (higher K_m values), but lower NADH K_m values. Despite a lowered k_{cat} , the R199A/R200A mutant exhibited a 200-fold coenzyme specificity switch towards NADH, although substrate inhibition was observed at high NADH concentrations ($K_i \approx 250 \mu M$). Stopped-flow FAD reduction studies confirmed substantially increased NADPH K_d values, although the limiting flavin reduction rate constant was similar in all mutants. The R199A mutation abolished electron transfer between hydroquinone FprA and NADP⁺, while this reaction progressed (via an FADH₂-NADP⁺ charge-transfer intermediate) for R200A FprA, albeit more slowly ($k_{lim} \approx 58.1 \text{ s}^{-1}$ compared with $>300 \text{ s}^{-1}$) than in WT. All mutations caused positive shifts in FAD potential (~ 40 – 65 mV). Binding of an NADPH analogue (tetrahydro-NADP) induced negative shifts in potential (~ 30 – 40 mV) only for variants with the R200A mutation, indicating distinctive effects of Arg₁₉₉/Arg₂₀₀ on coenzyme binding mode and FAD potential. Collectively, these data reveal important roles for the phylogenetically conserved arginines in controlling FprA FAD environment, thermodynamics, coenzyme selectivity and reactivity.

Key words: adrenodoxin reductase, coenzyme binding, electron transfer, enzyme mechanism, flavoprotein reductase A, *Mycobacterium tuberculosis*.